# Labeling index in pituitary adenomas evaluated by means of MIB-1: is there a prognostic role? A critical review

Paulo Henrique Pires de Aguiar<sup>1</sup>, Rogério Aires<sup>2</sup>, Edward R. Laws<sup>3</sup>, Gustavo Rassier Isolan<sup>4</sup>, Angela Logullo<sup>5</sup>, Chirag Patil<sup>6</sup>, Laurence Katznelson<sup>6</sup>

<sup>1</sup>Department of Neurology, São Paulo Medical School, State University of São Paulo, Brazil, <sup>2</sup>Divison of Neurosurgery, Santa Casa de Londrina Hospital, Brazil, <sup>3</sup>Department of Neurosurgery, Brigham and Women's Hospital, Boston, MA, USA, <sup>4</sup>Department of Neurosurgery and Postgraduation in Surgery, Federal University of Rio Grande do Sul, Porto Alegre, Brazil, <sup>5</sup>Department of Pathology, Federal University of São Paulo, Brazil, <sup>6</sup>Stanford Pituitary Center, Department of Neurosurgery, Stanford University, USA

Objective: The present article presents an overview of the literature, and analyses the methods and the primary questions related to assessment of proliferation index using the Ki-67/MIB-1 labeling index in pituitary adenomas. Although atypical adenomas are characterized by their atypical morphological features by an elevated mitotic index, a Ki-67 (MIB-1) labeling index greater than 3% and extensive nuclear staining for p53, use of the proliferation index (LI) of pituitary adenomas in assessing the degree of tumor aggressiveness is a controversial topic in the literature, and there are disparate results involving many studies

Methods: A review of literature was carried out to correlate the role of Ki-67 LI and its correlation with clinical findings, tumor size, invasiveness, recurrence, adenoma subtype, adenoma doubling time, and pituitary carcinomas is addressed.

**Results:** The prognosis cannot be predicted on the basis of the Ki-67 LI alone. Although there is no direct relation between Ki-67 LI and some of these variables and controversial data were found regarding some topics, our review justify the use of Ki-67 in the analysis of pituitary adenomas as an additional information for clinical decision.

Conclusion: Although assessment of proliferative may be helpful in predicting subsequent tumor recurrence or invasiveness, there are many other important and as yet unidentified factors pituitary tumors. It is clear that further research is needed to clarify these molecular mechanisms to predict those with a potentially poor clinical outcome.

Keywords: Pituitary adenomas, Labeling index, Cell growth, Invasion, MIB-1, Ki-67

#### Introduction

Anterior pituitary adenomas usually are histologically benign, with well defined behavior. However, in about one-third of cases, these adenomas infiltrate the surrounding tissues including the wall of the cavernous sinus. This local invasion adversely affects the surgical results and contributes to the possibility of relapse. 1–8

It is well accepted that brain tumor growth results from the relative proportion of cells contained in three populations: (1) cycling/proliferative; (2) quiescent (GO)/static; and (3) terminally differentiated/dying. Ki-67 expression is detected by the monoclonal antibody MIB-1 and is expressed as a

Correspondence to: Paulo Henrique Pires Aguiar, MD, PhD, Department of Neurology São Paulo Medical School, State University of São Paulo, São Paulo, Brazil. Email: phpaneurocir@gmail.com

percentage of immunopositive nuclei in the form of a Ki-67 LI. Generally, pituitary tumor with aggressively behavior has increased Ki-67 LI.

The cells in S phase (DNA duplication) were assessed in the past by means of autoradiographic analysis using [³H]thymidine, and in the 1980s with bromodeoxyuridine (intravenous infusion of 5-bromodeoxyuridine, BrdU, 200 mg/m²), to label tumor cells in the deoxyribonucleic acid (DNA) synthesis phase, S phase. The same autoradiographic principle, a flow cytometry method was also used for the same purpose. Using flow cytometry in 21 pituitary adenomas, Nagashima *et al.* showed that the S phase fraction was less than 0.1% in nine cases, 0.1–0.5% in seven, and greater than 0.5% in five. Except for two cases of Nelson's syndrome, in which it was greater than 1%, the S-phase fraction did not correlate with

any other variables, including patient age, tumor size, or duration of signs and symptoms. The small S phase fraction of most pituitary adenomas correlates well with the clinical behavior of these tumors, which grow much more slowly than other types of brain tumors such as gliomas. <sup>12</sup> The higher S phase fractions may reflect aggressive and invasive growth. These methods were abandoned because of potential side effects related to teratogenicity and myelosuppression.

The cycling compartment (G1, G2, M-mitosis, and S phases) can be detected by a mouse monoclonal Ki-67 antibody; an available, rapid, safe, sensitive, and specific method for immunostaining proliferative cells. 9,13-19 The monoclonal antibody Ki-67 reacts with a human nuclear cell proliferation-associated antigen that is expressed in all active parts of the cell cycle. Cattoretti et al. 20 raised monoclonal antibodies, MIB 1-3, against recombinant components of the Ki-67 antigen, and showed that these antibodies were true Ki-67 equivalents, as demonstrated by immunostain of fresh specimens, along with biochemical, and molecular biological techniques. These authors used formalin-fixed, paraffin-embedded sections routinely processed for immunohistochemistry in this study, antibodies MIB-1 and MIB-3 labeled mitotic figures, while non-mitotic proliferating cells did not label under these conditions. However, when dewaxed microwave oven-processed paraffin sections of formalin-fixed tissues were used, MIB-1 and MIB-3 produced strong nuclear staining of those cells presumed to proliferate under a variety of normal and neoplastic conditions. Moreover, routine decalcification or depigmentation techniques did not alter the immunoreactivity of MIB-1 and MIB-3 with microwave-processed paraffin sections.<sup>20,21</sup>

The Ki-67 monoclonal antibody has been used in many types of tumors, including tumors of the central nervous system (CNS)<sup>15,22,23</sup> and in skull base tumors. <sup>9,13,14,24–26</sup>

Burger *et al.* described assessment of the proliferation indices in pituitary adenomas in their analysis of proliferating cells in tumors of the CNS.<sup>22</sup> However, the first article that assessed the proliferation-associated Ki-67 in fresh-frozen pituitary adenoma specimen was published by Landolt *et al.*<sup>27</sup> Subsequently, extensive studies have attempted to correlate the Ki-67/MIB-1 labeling index (LI) with many clinical parameters and other biological markers.

LI may be defined as the proportion of labeled cells to the total number of cells analysed in a field consisting of between 100 and 1000 cells (arbitrary definition), in the area where the density of labeled cells is the highest. As a rule, the LI measured by Ki-67 shows an average rate of  $1.9 \pm 1.3\%$  in pituitary adenomas.

Beyond any doubt, the MIB-1 monoclonal antibody is one of the most important immunocytochemical markers for proliferation and its evaluation in routinely processed paraffin-embedded tissue specimens of pituitary adenomas. <sup>15–17</sup> Other factors such as the pituitary tumor transforming gene (PTTG), p16, p27, p53, vascular endothelial growth factor (VEGF), topoisomerase II-alpha, D1 cyclin, metalloproteinases, cadherin expression, cyclooxygenase, and aquaporin may also be studied, to be characterized in parallel with the MIB-1/Ki-67 LI. Together with the MIB-1/Ki-67 LI, these markers add information regarding the true biological role of cellular proliferation in a pituitary tumor.

Comparisons among the main methods that measure the LI were performed by Morimura *et al.*<sup>28</sup> This study showed that *in vitro* BrdU labeling can be a useful alternative to Ki-67 immunolabeling of human brain tumor specimens. Owing to collateral negative effects of BrdU, Ki-67 is the current choice for proliferation studies.<sup>28</sup>

The development of novel chemotherapy or gene transfer techniques is based on advances in our understanding of the genetic basis of pituitary adenoma invasiveness and proliferation. It is necessary to understand which factors may interfere with growth and its interruption, and which environmental factors might be involved in the genesis of these tumors. This is a comprehensive review of published data, serving as background for further studies.

The following topics are addressed in this review. We begin reviewing the aspects of immunocytochemical technique. Then we reviewed the relation of Ki-67 LI with clinical findings, tumor size, invasiveness, recurrence, adenoma subtype, adenoma doubling time, and pituitary carcinomas.

#### Aspects of the Immunocytochemical Technique

MIB-1 immunostaining is a simple and practical method that can be used in the routine histological evaluation of brain tumors and also of pituitary adenomas. Although automated analysis is faster and easier, manual cell counting is equally reliable and is applicable everywhere. A high growth fraction expressed by a high Ki-67 LI might suggest the need for careful clinical and radiological follow-up.<sup>29–31</sup>

Tumor cell proliferation has been assessed immunohistochemically by means of the MIB-1 antibody epitope of the Ki-67 LI antibody. The method may be used in paraffin-embedded specimens and in frozen sections. Microwave processing is necessary for paraffin sections, in order to promote deparaffination. Sections are cut at 4 µm. Antigen retrieval is obtained by boiling sections in citrate buffer. The chromogen used is a well-known immunohistochemical stain with avitin/biotin or peroxidase/anti-peroxidase (APAAP) diaminobenzidine. MIB-1 is apparent as nucleolar staining, related to fibrillar proteins in nucleoli, in the cells in the S, G1, G2, and M phase of the cell cycle. In each specimen, a total of at least 1000 tumor cell nuclei are evaluated. The count of

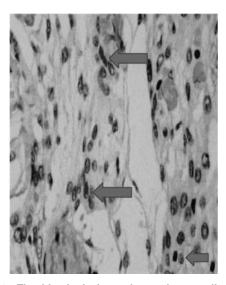


Figure 1 The histological specimen shows cells labeled immunohistochemically using the monoclonal antibody MIB-1, reaction APAAP (peroxidase anti-peroxidase), with diaminobenzidine as chromogen (dark brown nuclei), ×200. The red arrows show high density areas of labeled cells. Specimen obtained from a non-functioning pituitary adenoma, with suprasellar extension and invasion of the cavernous sinus. The estimated LI was 3% in the region of the highest density of labeled cells.

labeled cells obviously depends on interobserver variability and sampling error. <sup>15,16,20,30,31,32</sup> Because MIB-1 depends on tissue processing, staining method, and method of assessment, variability of results may be expected; however, results typically fall within a limited statistical spectrum. The ideal cut point for distinguishing the group of normal slow growth from faster growth is between 1.5 and 3%. <sup>33,34</sup> Figure 1 is placed in order to illustrate the concept and practical application of the technique.

Mastronardi *et al.*<sup>35</sup> used the classical method<sup>9,20,21</sup> using avidin–biotin–peroxidase. Ten fields were selected in regions with the highest concentrations of MIB-1-positive nuclei and were examined high power (×400). Each field corresponded to a total number of cells ranging from 700 to 1000. Areas of necrosis, normal adenohypophysial cells, and endothelial cells were excluded from the evaluation. Considering 1000 cells with 'manual' counting, the Ki-67 LI has been defined as the percentage of MIB-1 positive cells (dense brown precipitate restricted to the nuclei). In our opinion, this is the standard method to be followed.

# Is There Any Correlation between the Clinical Course of Pituitary Adenoma Patients and the Ki-67/MIB-1 LI?

Age

As described by the majority of authors in the reviewed literature, <sup>36–39</sup> there is an inverse correlation between Ki-67 LI and age for non-functioning adenomas. However, this finding is not described in other studies. <sup>40–42</sup>

#### Gender

Several studies showed no correlation between Ki-67/MIB-1 LI and gender. However, Delgrande *et al.* demonstrated statistically higher LI in males compared to females with large prolactinomas. Paek *et al.* and Qian *et al.* confirmed the studies of Delgrande *et al.* Paek *et al.* found similar results in microadenomas.

A predominance of higher LI in females was found by Wolfsberger et al. in 2004 regarding non-functioning pituitary adenomas. 45 Schaller 47 studied growth hormone (GH) secreting adenomas from 18 patients (10 males) with acromegaly who met strict immunohistochemical and electron microscopic diagnostic criteria and who underwent surgical resection of their tumors. The men and women were equivalent in age at surgery.<sup>35</sup> Men demonstrated higher insulin-like growth factor-1 (IGF-1) and lower GH levels preand post-operatively, whereas the percentage reduction in IGF-1 was more pronounced in men when compared to women (58% versus 27%). The overall outcome was better in women than in men.<sup>47</sup> The mitosis and MIB-1 LIs were increased in men compared to women. Finally, this article concluded that clinical course and tumor biology of GH-releasing pituitary adenomas appeared to differ between women and men. Therefore, the correlation between LI and gender is unclear for pituitary adenomas.

#### Signs and symptoms

Ki-67 LI results have been shown to correlate with signs and symptoms related to the pituitary adenomas. According to Shaller, shorter pre-operative duration of symptoms, larger and more invasive tumors, and a worse clinical outcome were associated with higher Ki67 LI.<sup>47</sup> Paek *et al.* determined that visual defects correlated with higher Ki-67 LI in patients with macroadenomas.<sup>39</sup>

Suzuki *et al.* studied Ki-67 LI in 52 incidentally detected pituitary adenomas found lower rates of LI in tumors of non-symptomatic patients than in symptomatic patients. <sup>42</sup> Finally, Matha *et al.* found an association between Ki-67 LI and unilateral temporal hemianopsia in patients with non-functioning adenomas. <sup>46</sup> These studies suggest that the Li does correlate with tumor aggression as based on signs of mass effects and symptoms.

#### Pre-operative medical therapy

The Ki-67 LI is lower in octreotide-treated GH and bromocriptine-treated prolactin adenomas, suggesting that these drugs probably inhibit growth in pituitary adenomas. The other studies have not demonstrated a statistical difference in Ki-67 LI between pre-operative drug-treated patients and untreated patients.

#### Surgical treatment

There are limited data in this regard. Losa et al.<sup>50</sup> reported that a higher LI was demonstrated in

Cushing's adenomas patients not cured by surgery compared with tumors from patients in surgical remission, though better correlations occurred with maximal tumor diameter and basal ACTH levels.

#### Radiation therapy

Radiation therapy has an important role in the therapy of pituitary adenoma, but it is usually indicated as adjuvant therapy, and case selection is a controversial topic in pituitary tumor therapy. This controversy includes a debate concerning the choice of conventional fractionated therapy versus stereotactic radiation therapy. 51,52

In order to elucidate histological changes in the pituitary gland and adenomas following radiotherapy, two irradiated pituitary glands and seven irradiated non-functioning adenomas were studied by Nishioka *et al.*, and it was observed that in pituitary adenomas, the MIB-1 LI remained unchanged after radiation.<sup>53</sup> The histological changes were more intense in adenomas following gamma knife radiotherapy than those following conventional radiotherapy alone, though the true impact of stereotactic radiotherapy versus conventional radiotherapy on the labeling index in pituitary adenomas is largely unknown.

#### Ki-67 Expression/MIB-1 and Tumor Size

Because of sellar confinement of pituitary adenomas, growth velocity may be reduced or inhibited in a subset of macroadenomas. Microadenomas have no

restriction to growth in the sellar space and initially may have a high growth velocity, reaching a higher LI index than noted in many macroadenomas. This idea is reinforced by the study by Yonezawa *et al.* <sup>38</sup> As described by Turner *et al.* <sup>54</sup> Ki-67 LI share significantly higher in macroprolactionomas than in microprolactinomas. This finding was not corroborated by others. <sup>35,43</sup>

In contrast, ACTH secretion correlates with the size of the corticotroph tumor and a high rate of MIB-1 labeling,<sup>50</sup> perhaps because the functional adenomas have their growth influenced by many other factors, and contact inhibition may be not so important. It suggests that tumor subtype is important when considering Ki-67 staining and tumor size.

### Ki-67 Expression/MIB-1 LI and Invasiveness. Is There a Valid Correlation?

A tendency for parasellar expansion of many anterior pituitary adenomas has well been described in the literature. 55–58 Jefferson identified a group of pituitary tumors classified as 'invasive' adenomas, in which extrasellar and parasellar spread might occur. 56 An invasive pituitary adenoma was defined as a tumor that extended beyond its capsule or involved contiguous structures. The incidence of invasiveness among these tumors varies among different anterior pituitary adenoma subtypes 56,59 and also in relation to the criteria used for assessment. Table 1 summarizes the main studies correlating

Table 1 Literature review of the correlation between Ki-67/MIB-1 LI and Invasiveness in pituitary adenomas

Reference	n	GH	PRL	ACTH	TSH	NF	FSH and/or LH	Plurihormonal	Association with tumor invasiveness
64	32					32			No correlation
83	25		25						↑ Increased ↑ in invasive*
46	85					85			Increased in invasive
95	213					213			No correlation
65	35	2	7	1		1	5	19	↑ Increased in invasive case ↑ **
39	44	8	8			28			Negative correlation
26	1					1			Increased in invasive case
81	65	21	6	6	2	2	28		↑ Increased ↑ in invasive
66	159	43	19	16		42		39	↑ Increased in invasive**
45	260 (MIB-1)	34	64	36	4	67	39	16	† Increased ↑ in invasive**
68	26 (Ki-67)	8	4	4		9		1	↑ Increased ↑ in invasive**
34	23					23			No correlation
37	132	42	15	6	1	68			↑ Increased ↑ in invasive***
82	69	7	9	4	1	37	5	6	↑?Increased in invasive**
35	121	24	26	12	3	48	1	7	↑ Increased in invasive**
71	94	18	20	5		41		10	No correlation
62	15	15							↑ Increased in invasive*
54	160	46	29	18		67			Negative correlation
41	103	21	24	10		41		7	↑ Increased ↑ in invasive**
69	123	34	15	6		68			↑ Increased ↑ in invasive**
70	48								↑ Increased in invasive*
38	85	11	17			57			↑ Increased in invasive
40	45	9	8	1		27			No correlation
43	96		96						No Correlation
4	31	4	10	3		12	2		↑ Increased ↑ in invasive**
30	127	23	8	7	3	73		6	↑ Increased in invasive*
49	65	10	20	7	3	24			No correlation

Note: n = no. of cases; GH = growth hormone adenoma; PRL = prolactin adenoma; ACTH = adenocorticotropin hormone adenoma; TSH = thyroid simulating hormone adenoma; FSH = follicle stimulating hormone adenoma; LH = luteinizing hormone adenoma; plurihormonal = adenoma with two or more hormones; NF = non-functioning adenoma;  $\uparrow$  = higher; \*P < 0.005; \*\*P < 0.05; \*\*\*not statistically significant.

invasiveness and proliferative index assessed by MIB-1/Ki-67 LI.

The most accepted criterion to classify invasiveness is radiological, by means of coronal sections of the MRI as suggested by Knosp et al.<sup>6</sup> The grades 0–3 are distinguished from each other by a medial tangent, the intercarotid line-through the cross-sectional centers, and a lateral tangent based on the intra- and supracavernous internal carotid arteries.<sup>6</sup> Grade 0 is the tumor which does not invade the cavernous sinus, and Grade 4 corresponds to total encasement of the intracavernous carotid artery. In a series of 25 surgically-treated invasive pituitary adenomas, Knosp et al.6 used this classification and found surgically proven invasion of the cavernous sinus space present in all Grades 3 and 4 cases and in all but one of the Grade 2 cases; no invasion was present in Grades 0 and 1 cases. The critical area where invasion of the cavernous sinus space becomes very likely and can be proven surgically is located between the intercarotid line and the lateral tangent, which is represented as Grade 2. In this article, Knosp et al.<sup>6</sup> also measured tumor growth rates, using the monoclonal antibody Ki-67, and showed a statistically higher proliferation rate (P < 0.001) in adenomas with surgically observed invasion into the cavernous sinus space, as compared with non-invasive adenomas.

In macroadenomas of the pituitary gland, invasion of parasellar spaces may occur in 6–10% of cases. With high-resolution high-field (3 T) MRI, the sellar region may be evaluated more accurately as compared to lower field strengths. 60

Many authors have shown a correlation of Ki-67/MIB-1 LI with tumor growth velocity<sup>5,34,37,61</sup> and tumor invasiveness into skull base anatomically adjacent structures.<sup>4,26,30,36,41,61–69</sup> Thapar *et al.*<sup>30</sup> established a Ki-67 LI threshold of 3% for distinguishing between non-invasive and invasive adenomas, with specificity and sensitivity of 97 and 73%, respectively. Mizoue *et al.* suggested a cut point of 1% for non-invasive versus invasive tumors. This series included eight adenomas with rapid growth out of a total of 48 non-invasive tumors.<sup>70</sup>

Yonezawa *et al.*<sup>38</sup> found no difference in Ki-67 Li between invasive and non-invasive functioning microadenomas. In a study of 103 pituitary adenomas, Mastronardi *et al.*<sup>35</sup> concluded that the cut points were 3.5% for invasive adenomas and 5% for cavernous sinus involvement, much higher than proposed by Thapar *et al.*<sup>30</sup> The mean cut point, however, could not be used in all series, because, in some series, the LIs in invasive tumors were above 3 or 3.5%.<sup>69</sup> Zhao *et al.* found a mean LI of 0.75% for non-invasive adenomas compared to 2.2% in invasive tumors.<sup>69</sup>

Mastronardi *et al.*<sup>41</sup> studied 24 microadenomas, 27 intrasellar macroadenomas, 34 intra-suprasellar macroadenomas, and 36 intra-supra-parasellar

macroadenomas. In their analysis, there were 76 non-infiltrating adenomas and 45 infiltrating adenomas. The wall of the cavernous sinus (CS) was infiltrated in 18 cases. Forty-eight adenomas were non-functioning and 73 were functioning. The LIs were  $3.73 \pm 5.13\%$  in infiltrating and  $2.03 \pm 2.41\%$  in non-infiltrating adenomas (P = 0.02), and  $5.61 \pm 7.19\%$  in CS-infiltrating versus  $2.09 \pm 2.37\%$  in CS-non-infiltrating adenomas (P = 0.0005). This study demonstrated a higher LI in the more aggressive, infiltrating tumors.

Lath *et al.*<sup>71</sup> demonstrated that the LI correlated with clinical and radiological evidence of invasiveness. These authors demonstrated that the mean Ki-67 LI for all pituitary adenomas was 0.84% (range: 0–17.45%) and that voluminous invasive macroadenomas (1.44%) had a higher Ki-67 LI as compared to microadenomas (0.36%). The authors concluded that the difference in the Ki-67 LI between invasive and non-invasive adenomas was not statistically significant; hence, for their series of 94 pituitary adenomas, the Ki-67 LI was not a reliable indicator of invasiveness in pituitary adenomas.<sup>71</sup>

The topic is so controversial that Pizarro *et al.*<sup>66</sup>, in a well performed study involving microadenomas, found significantly higher Ki-67 LI in invasive than non-invasive tumors. However, a well-defined LI cut point was not established due to overlap in the LI between the two groups. Paek *et al.*<sup>39</sup>, in their study of pituitary macroadenomas, did not detect a significant difference in LI between invasive and non-invasive, as well as between non-functioning and functioning tumors.<sup>39</sup> Mahta *et al.*<sup>72</sup> studied 85 non-functioning pituitary adenomas and did not find a correlation between Ki-67 LI with either invasiveness or recurrence.

In the most recent World Health Organization Classification of Tumors of Endocrine Organs (2007), Ki-67 LI higher than 3% defines a pituitary adenoma as atypical, but based on the literature addressed above, this cutoff of 3% is controversial and, besides, even for some authors, <sup>39,72</sup> there is no relation between invasiveness and Ki-67.

## Ki-67 Expression/MIB-1 and Recurrence: Important for Prognosis?

Because Ki-67 LI correlates with tumor proliferation, it has been suggested that Ki-67 may correlate with risk of recurrence. Abe *et al.*<sup>40</sup> demonstrated a recurrence rate of 50% in tumors with Ki-67 LI higher than 1.5% and a recurrence rate of 16% in tumors with LI <1%. Turner *et al.*, in non-functioning adenomas, demonstrated no correlation between Ki-67 LI and recurrence, concluding that this marker has no utility as a predictor of tumor behavior.<sup>54</sup> Nakabayashi *et al.*<sup>72</sup> emphasized that subtotal surgical resection is an important factor contributing to recurrence. In this study, the Ki-67 LI and analysis

1064

of cyclin A rates were evaluated and found to be predictive of shorter progression-free survival.<sup>72</sup>

In recurring non-functioning pituitary adenomas (NFPAs), the initial MIB LI after surgery has been shown to be statistically higher than in tumors with no regrowth.<sup>5</sup> Though this topic is controversial, Honegger *et al.*<sup>34</sup> showed a significant correlation between MIB-1 LI and velocity of regrowth, which was confirmed by Tanaka *et al.*<sup>36</sup> No statistical difference regarding NFPAs has been observed by other authors.<sup>37,50,73</sup>

Some studies suggest that the regrowth potential in the absence of postoperative radiotherapy or radiosurgery may reach a rate between 38 and 95%; however, lack of consistent follow-up methodology may create a bias in the analysis. 74–78 Recurrence may be present in the 3–6 months following surgery, as detected mainly by T1 MRI in the coronal section images, with gadolinium.

However, it is important to keep in mind that some studies showed that residual tumors may remain unchanged or have minimal growth in almost 48% for a long period of time.<sup>75,76</sup>

In another study, Ki-67/MIB-1 was a better predictor than PTTG as a marker of recurrence, particularly with follow-up greater than 1 year. <sup>79</sup> The cut point of 2.9% for Ki67 for pituitary adenomas showed a higher incidence of recurrence. <sup>79</sup> In a series of 176 pituitary tumors, Scheithauer *et al.* <sup>48</sup> found no correlation of Ki-67 LI with recurrence based on follow-up of 78 patients, of whom only seven had indices higher than 3%.

The most recent World Health Organization Tumor Classification of Tumors of Endocrine Organs defines an atypical pituitary adenoma as a tumor with Ki-67 LI higher than 3%. The controversy of this definition is that these tumors are uncommon and that there is aggressive behavior in tumors with Ki-67 LI lower than 3%.

Therefore, the literature suggests that use of Ki-67 LI alone has limited prognostic ability to predict recurrence accurately, and the association with other biological markers may be more promising. The main studies in literature are summarized in Table 2.

MIB-1 LI and pituitary adenoma doubling time Several authors have found an inverse correlation between Ki-67 LI and tumor volume doubling time which was statistically significant. These data support a role for Ki-67 LI to predict tumor aggression.

#### MIB-1 LI and pituitary adenoma subtypes

It has been demonstrated that Ki-67 staining was higher in functioning tumors than in non-functioning tumors. <sup>6,24,30,37,41,42,48,68</sup> Some articles, <sup>40,69,80–82</sup> such as those published by Paek *et al.*, showed no correlation between Ki-67 LI and type of pituitary adenomas. <sup>39</sup> Table 3 summarizes the main studies correlating MIB-1/Ki-67 LI and pituitary adenoma subtypes.

#### Prolactin (PRL) secreting tumors

Asano *et al.*<sup>1</sup> evaluated the MIB-1 index in 63 surgically removed pituitary adenomas; values ranged from 0 to 6.5%, with lower values in growth hormone secreting tumors and high values in prolactin secreting tumors. Pre-operative treatment with bromocriptine had no effect on the LI values.<sup>1</sup> Mastronardi *et al.*<sup>41</sup> also found that growth hormone secreting adenomas had a low Ki-67 LI, whereas prolactin secreting tumors had a mean index above the mean for the cohort as a whole. Delgrande *et al.*<sup>43</sup> demonstrated that males harboring large macroprolactinomas have a higher LI as compared with females with PRL adenomas. Others authors reported similar results.<sup>39,44</sup>

Wierinckx et al.<sup>83</sup> showed that markers of proliferation could not differentiate invasive from non-invasive PRL secreting adenomas. This study

Table 2 Correlation between MIB-1/Ki-67 and recurrence of pituitary adenomas

Reference	n	GH	PRL	ACTH	TSH	NF	FSH and/or LH	Plurihormonal	Correlation with tumor recurrence
46	85					85			No correlation
87	32						32		No correlation
83	25		25						↑ No correlation
79	45	8	14	6		6	11		in recurrent tumors*
31	176	20	35	18	10	50	20		No correlation
39	44	8	8			28			> recurrent tumors*
66	159	43	19	16		42		39	No correlation
34	23					23			> recurrent tumors
36	40					40			> recurrent tumors
73	51								No correlation
37	132	42	15	6	1	68			No Correlation
72	48								< progression free interval**
50	101					101			No Correlation
94	67					67			No Correlation
5	33					33			Increased velocity of growth in recurrence
49	65	10	20	7	3	24			Increased in recurrence**

Note: n = total; GH = growth hormone adenoma; PRL = prolactin adenoma; ACTH = adenocorticotropin hormone adenoma; TSH = thyroid simulating hormone adenoma; FSH = follicle stimulating hormone adenoma; LH = luteinizing hormone adenoma; Pluri = adenoma with two or more hormones; NF = non-functioning adenoma; > = higher; < = lower; \*P < 0.05; \*\*P < 0.005.

evaluated mitotic activity, Ki-67 LI, and p53 labeling, and demonstrated that mitotic index and Ki67 LI were elevated in five of 25 invasive tumors.

These studies suggest that LI as measured by Ki-67 may be, in part, dependent on the type of pituitary adenoma.

#### ACTH secreting tumors

Several authors have found a high Ki-67/MIB-1 in ACTH secreting tumors, with values similar to recurrent adenomas and non functioning adenomas. <sup>24,30,35,49</sup> Katznelson *et al.* <sup>84</sup> reported higher Ki-67 LI in macroadenomas producing ACTH (44% of macroadenomas with high LI) than microadenomas (only 18% of microadenomas with higher LI). Losa *et al.* <sup>61</sup> also reported that a high LI was demonstrated in Cushing's adenoma patients not in remission in the post-operative period when compared with patients in remission. Significant correlations were present with maximal tumor diameter and basal ACTH levels, <sup>61</sup> with results similar to those of Katznelson *et al.* <sup>84</sup>

Several authors have reported a higher Ki-67 LI in ACTH secreting tumors than other functioning secreting tumors even when ACTH microadenomas are included.<sup>35,49,85</sup> Some 'silent' corticotroph tumors may have the potential for ACTH secretion leading to hypercortisolemia at a later stage in the disease but Ki-67 does not predict this behavior.

#### GH secreting tumors

Shaller,<sup>47</sup> studying 18 GH secreting tumors by means of MIB-1 and mitosis index, suggested that therapy for GH-releasing adenomas should be more

aggressive in men than in women. The gender-related differences in GH-releasing pituitary adenomas appear to have a basis in different biologic behavior, suggesting that the LI and aggressive invasion might be higher in men.<sup>47</sup>

GHRH mRNA was correlated with high rates of Ki-67 LI demonstrating that the aggressiveness of tumors was related to GHRH-mRNA.<sup>86</sup>

#### Gonadotrophin secreting tumors

There are limited data available for these tumors. Dubois *et al.*<sup>87</sup> found no correlation between Ki-67 LI and residual tumor, as well as ki-67 LI and recurrence in 32 gonadotropic pituitary adenomas. They did find a correlation of recurrence with age and antero posterior diameter.<sup>87</sup>

#### Silent and null cells adenomas

Clinically non-functioning pituitary adenomas are classified by Asa and Kovacs in two groups. The first group is related to those tumors which have hormone immunoreactivity and ultrastructural features of known adenohypophyseal cell types. The former involves the silent somatotroph adenomas, silent corticotroph adenomas, and silent gonadotroph adenomas. The second group includes silent type III adenomas, null cell adenomas, and oncocytomas. Silent corticotroph adenomas are rare tumors defined as pituitary tumors with ACTH immunoreactivity, but without clinical evidence of Cushing's disease.

The growth characteristics of clinically silent pituitary adenomas are poorly understood. Therefore, the radiologically measured growth of

Table 3 Correlation between KI-67/MIB-1 LI and pituitary adenoma subtype

Reference	n	GH	PRL	ACTH	TSH	NF	FSH and/or LH	Pluri	Association with tumor subtype or functional status
31	176	20	35	18	10	50	20		> in HS than NF*
42	95+9 silent					43S	22		> in NF than incidentalomas**
	+22 gonadotrophic adenomas					43N S			
80	38	6	9					23	No statistical correlation
39	44	8	8			28			No statistical correlation
81	65	21	5	3	3	34			No statistical correlation
66	159	43	19	16		42		39	No statistical correlation
45	260	34	64	36	4	67	39		> in HS than NF**
37	132	42	15	6	1	68			> in HS than NF*
82	69	7	9	4	1	37	5	6	No statistical correlation
35	121	24	26	12	3	48	1	7	No statistical correlation
61	51			51–36					> Ki-67 LI in macroadenomas than
				macro- and 15 microadenomas					microadenomas**
69	123	34	15	6		68			No statistical correlation
41	103	21	24	10		41		7	> in ACTH than other HS tumor**
84	33	_ '	2-7	33–16		7.		,	> Ki-67 LI in macroadenomas than
04	00			macro- and 17 microadenomas					microadenomas
40	45	9	8	1		27			No statistical correlation
30	127	23	8	7	3	73		6	> in HS than NF**
39	65	10	20	7	3	24			> in HS than NF**

Note: n = total; GH = growth hormone adenoma; PRL = prolactin adenoma; ACTH = adenocorticotropin hormone adenoma; TSH = thyroid simulating hormone adenoma; FSH = follicle stimulating hormone adenoma; LH = luteinizing hormone adenoma; Pluri = adenoma with two or more hormones; NF=non-functioning adenoma; HS = hormone secreting tumors, S = symptomatic; NS = non-symptomatic, LI = labeling index; > = higher; \*\*P < 0.05; \*statistically not significant.

inactive pituitary adenomas may be analysed and compared with adenoma classification and immunostaining for proliferation markers.<sup>64</sup> In a series of 32 patients with non-functioning pituitary adenomas who underwent 45 operations, Saeger et al. 64 analysed the correlation between invasiveness by means of MRI and immunostained characteristics (PCNA, MIB-1, P53 and IGF-1), and concluded that statistically significant differences were present for growth rate and PCNA expression. P53 immunostained positive in invasive adenomas only.<sup>64</sup> There were no correlations with clinical growth rate, but p53 expression correlated significantly to numbers of MiB-1-positive nuclei and PCNA-positive nuclei. The mean LI for MiB-1 was 0.12 in adenomas growing less than 1.5 mm per year and 0.34 in adenomas growing more than 1.5 mm per year. For non-invasive adenomas, the MiB-1 LI was 0.03, for invasive adenomas, it was 0.126, and for strongly invasive adenomas, it was 0.212. The MiB-1 LI was lower in null cell adenomas than in FSH/LH adenomas. Otherwise, all these data for MIB-1 showed no statistically significant differences (P <0.05). This study found that the PCNA LI in adenomas growing less than 1.5 mm per year was 0.51 in contrast to LI of 1.12 for those growing more than 1.5 mm. In non-invasive adenomas, the PCNA

LI was 0.796, in invasive adenomas, it was 0.655 and in diffusely invasive lesions, it was 1.011. Finally, these authors recommend the use of PCNA if correlations with progression of tumor growth are desired.<sup>64</sup>

#### **Oncocytomas**

In one study, there was no correlation between oncocytic transformation and Ki-67/MIB-1 LI.<sup>81</sup> Because of the rarity of this transformation, further studies are necessary to achieve a definitive conclusion

#### Non-functioning adenomas

Pituitary adenomas without clinically active hypersecretion are summarized under the term non-functioning pituitary adenoma.

Complete surgical removal of non-functioning adenomas has been reported in up to 16–79% of cases. <sup>34,50,74–78,88–95</sup> This complicates the evaluation of the influence of MIB-1 on recurrence, because depending on grade of resection, the remnant tumor volume may have a greater apparent regrowth potential.

Yonezawa *et al.*<sup>38</sup> showed higher Ki-67 LI in non-functioning microadenomas compared to expansive and invasive adenomas, regardless of function. Turner *et al.*<sup>54,94</sup> have reported no correlation between Ki-67 LI and recurrence in non functioning adenomas.

Table 4 The correlation between biological markers and Ki-67/MIB-1 in pituitary adenomas

Reference	n	Marker	Conclusion						
106	73	Metaloproteinase MMP-9	Increased in invasion and Ki-67						
64	32 (non-functioning adenomas)	P53	Correlation with PCNA and MIB-1*						
83	25	PTTG P53	Indirect correlation with invasiveness and recurrence						
95	213	E-cadherin MMP-9 P53 Ptd-FGFR4	No correlation in cavernous sinus pituitary adenoma invasion						
42	52 (incidentalomas)	D-Topoisomerase	No correlation						
79	45	PTTĠ	Correlation with Ki-67						
80	38	c-erb2	Variable positivity for Ki-67						
105	54	Metaloproteinase MMP-2	No correlation with Ki-67						
104	117	c-erb B2	No correlation with Ki-67						
65	35	MVD (F8) VEGF	Indirect correlation in invasive mainly MVD and VEGF						
		c-Myc bcl 2	MMP-9*						
		MMP-9	No correlation c-myc,bcl-2						
45	260	D topoisomerase II Alpha	Increased in higher rates of MIB-1**						
44	39 (13 invasive, and 26 non-invasive)	E-cadherin/beta cadherin	Increased in tumors with high rates of Ki-67						
82	69	P53/Mdm2	Increased in invasive tumors and Ki-67 increased in invasive tumors						
72	48	A ciclin/p27	No correlation						
54	160	Bcl 2, MVD	Increased in invasive tumours, and Ki-67 also increased						
94	67	Cyclin A, B, D, and E	Cyclin A, B, D and E higher in macroadenomas than microadenomas. In NF adenomas higher cyclin D and Ki-67						
62	25(secreting GH)	VEGF	Increased and Ki-67 increased in invasive tumors						
69	123	P27	No correlation						
86	91 (secreting GH)	GHRAmRNA	Linearly correlation with Ki-67 LI						
30	12 (invasive)	P53	P53 and Ki-67 higher in invasive adenomas						

Note: MMP = metaloproteinases; PTTG = pituitary transformer tumor gene; Mdm = murine double minute; PCNA = proliferative cell nuclear antigen minute; VEGF = vascular endothelial growth factor; MVD = microvessel density; NF = non-functioning; GHRH-mRNA = gene releasing hormone receptor; ptd-FGFR4 = pituitary tumor-derived fibroblast growth fraction receptor 4; \*statistic significance; \*\*statistically not significant.

Suzuki *et al.*<sup>42</sup> demonstrated that Ki-67 LI was higher in non-functioning tumor than secreting adenomas in a group of small tumors.

It is necessary to mention that most 'non-functioning adenomas' are in fact gonadotroph adenomas.

#### Incidentalomas

Some authors found lower values of LI values in incidentally discovered pituitary masses, mostly from autopsy studies, than pituitary adenomas in symptomatic patients. In a study of eight incidental pituitary microadenomas identified from 120 pituitary glands procured from MIB-1, staining for the Ki-67 LI antigen showed absolutely no expression. <sup>96</sup> The low proliferation indices in these tumors may reflect the small tumor size. <sup>42,96</sup>

The correlation of others biomarkers (PTTG, securin, H-ras, c-Myc, c-erb, P53, nm23, Rb genes, bcl-2, cyclin, p27, cadherins and catenins, and metalloproteinases) with Ki-67 LI in pituitary adenomas can give us more information regarding biological behavior of these tumors, 44,49,54,68,72,75,79,82,89,97,98,100,101–106 but is not the purpose of this article discuss these topics. Table 4 summarizes the main studies concerning Ki67/MIB-1 LI and their correlations with other biological markers for pituitary adenomas.

#### Pituitary carcinoma

These tumors are defined by the presence of brain, subarachnoid, or systemic metastasis. Pituitary carcinomas are rare and, although there is a higher Ki-67 LI (range: 7.8–11.91%), a progression of an adenoma-to-carcinoma sequence is not totally established: *de novo* pituitary tumorigenesis or a separate clonal expansion from the original tumor should be considered.

#### **Conclusions**

Currently, the main method for evaluating proliferation index in pituitary adenomas is immunohystochemical analysis by means of the monoclonal antibody MIB-1 to the Ki-67 LI antigen found in the protein of nucleoli of proliferating cells. Ki-67 can be measured in archived paraffin tissues or in frozen sections. There are also many other bio-molecular markers and genetic factors that may be important in understanding proliferation and invasiveness of pituitary adenomas. Further studies are necessary to determine the value of these markers in predicting tumor aggressiveness, and whether individual markers may have greater specificity with pituitary tumor subtypes.

Tumor size, invasion, and the incomplete tumor resection are important risk factors for recurrence or progression of pituitary adenomas.<sup>28</sup> Patient with progressive residual tumors showed invasiveness significantly more frequent than stable tumors.<sup>107</sup> The next investigations should find a Ki-67 LI

threshold for post-surgical tumor resection to define more precisely information regarding tumor progression. There is no evidence that an adenoma with higher Ki-67 LI will progress to carcinoma.

This review justifies the use of Ki-67 LI in the analysis of pituitary adenomas as a piece of additional information for clinical decision making because a high Ki-67 LI proliferative index in a pituitary adenoma might indicate a more aggressive behavior. We believe that the Ki-67/MIB-1 LI represents an additional piece of information that is helpful for clinical decision making. An adenoma with a high Ki-67 LI has an increased risk of early recurrence and invasiveness, and may need closer follow-up and aggressive adjuvant therapy. However, the overlap of Ki-67/MIB-1 LI, particularly in those adenomas with moderate growth velocity, suggests that the prognosis cannot be predicted on the basis of the Ki-67/MIB-1 LI alone.

#### References

- 1 Asano K, Kubo O, Tajika Y, Huang MC, Takakura K. The relationship between cell proliferation and secretory activity in pituitary adenomas. A review of 63 cases. No To Shinkei. 1996; 48: 543–49.
- 2 Atkin SL, Green VL, Hipkin LJ, Landolt AM, Foy PM, Jeffreys RV, *et al.*. A comparison of proliferation indices in human anterior pituitary adenomas using formalin-fixed tissue and in vitro cell culture. J Neurosurg. 1997; 87: 85–88.
- 3 Buchfelder M, Fahlbusch R, Adams EF, Kiesenwetter F, Thiereauf P. Proliferation parameters for pituitary adenomas. Acta Neurochir Suppl (Wien). 1996; 65: 18–21.
- 4 Daita G, Yonemasu Y. Dural invasion and proliferative potential of pituitary adenomas. Neurol Med Chir (Tokyo). 1996; 36: 211–14.
- 5 Ekramullah SM, Saitoh Y, Arita N, Ohnishi T, Hayakawa T. The correlation of Ki-67 staining indices with tumor doubling times in regrowing non-functioning pituitary adenomas. Acta Neurochir (Wien). 1996; 138: 1449–55.
- 6 Knosp E, Kitz K, Perneczky A. Proliferation activity in pituitary adenomas: measurement by monoconal antibody Ki-67. Neurosurgery. 1989; 25: 927–30.
- 7 Martins AN, Hayes GJ, Kempe LG. Invasive pituitary adenomas. J Neurosurg. 1989; 22: 268–76.
- 8 Pernicone PJ, Scheithauer BW. Invasive pituitary adenomas and pituitary carcinomas. In: Lloyd RV (ed.). Surgical pathology of the pituitary gland. Philadelphia, PA: WB Saunders; 1993. pp. 121–36.
- 9 Tsanaclis AM, Robert F, Michaud J, Brem S. The cycling pool of cells within human brain tumors: *in situ* cytokinetics using the monoclonal antibody Ki-67. Can J Neurol Sci. 1991; 18: 12–17.
- 10 Hoshino T, Rodriguez LA, Cho KG, Lee KS, Wilson CB, Edwards MS, et al. Prognostic implications of the proliferative potential of low-grade astrocytomas. J Neurosurg. 1988; 69: 839–42
- 11 Cho KG, Nagashima T, Barnwell S, Hoshino T. Flow cytometric determination of modal DNA population in relation to proliferative potential of human intracranial neoplasms. J Neurosurg. 1988; 69: 588–92.
- 12 Nagashima T, Murovic JA, Hoshino T, Wilson CB, de Armond SJ. The proliferative potential of human pituitary tumors *in situ*. J Neurosurg. 1986; 4: 588–93.
- 13 Aguiar PH, Tatagiba, M, Timpe ED, Matthies C, Samii M, Ostertag H. Proliferative activity of acoustic neurilemomas without neurofibromatosis determined by monoclonal antibody MBI 1. Acta Neurochirur (Wien). 1995; 134: 35–39.
- 14 Aguiar PH, Tsanaclis AM, Tella Junior OI, Plese JPP. Proliferation rate intracranial meningiomas as defined by the monoclonal antibody MIB-1: correlation with peritumoral oedema and other clinicoradiological and histological characteristics. Neurosurg Rev. 2003; 26: 221–28.
- 15 Gerdes J, Schwab Ū, Lemke H, Stein H. Production of a mouse monoclonal antibody reactive with a human nuclear

- antigen associated with cell proliferation. Int J Cancer. 1983; 31: 13–20.
- 16 Gerdes J, Dallenbach F, Lennert K, Lemke H, Stein H. Growth fractions in malignant non-Hodgkins lymphomas (NHL) as determined *in situ* with monoclonal antibody Ki-67. Hematol Oncol. 1984; 2: 365–71.
- 17 Gerdes J, Stein H, Pileri S, Rivano MT, Gobbi M, Ralfkiaer E, et al. Prognostic relevance of tumor-cell growth fraction in malignant non-Hodgkin's lymphomas. Lancet. 1987; 2: 448–49.
- 18 Gerdes J. Determination of the growth fraction by means of immunostaining with monoclonal antibody Ki-67. Acta Histochem Suppl. 1988; 36: 437–46.
- 19 Gerdes J, Becker MH, Key G, Cattoretti G. Immunohistological detection of tumor growth fraction (Ki-67 antigen) in formalin-fixed and routinely processed tissues. J Pathol. 1992; 168: 85–86.
- 20 Cattoretti G, Becker MH, Key G, Duchrow M, Schwater C, Galle J, et al. Monoclonal antibodies against recombinant parts of the Ki-67 antigen (MIB 1 and MIB 3) detect proliferating cells in microwave-processed formalin-fixed paraffin sections. J Pathol. 1992; 168: 357–363.
- 21 McCormick D, Chong H, Hobbs C. Detection of the Ki-67antigen in fixed and wax-embedded sections with the monoclonal antibody MIB-1. Histopathology. 1993; 22: 355– 60.
- 22 Burger PC, Shibata T, Kleihues P. The use of the monoclonal antibody Ki-67 in the identification of proliferating cells: application to surgical neuropathology. Am J Surg Pathol. 1986; 10: 611–17.
- 23 Vankalakunti M, Vasishta RK, Das Radotra B, Khosla VK. MIB-1 immunolabeling: a valuable marker in prediction of benign recurring meningiomas. Neuropathology. 2007; 27: 407–12.
- 24 Knosp E, Steiner E, Kitz K, Matula C. Pituitary adenomas with invasion of the cavernous sinus space: a magnetic resonance imaging classification compared with surgical findings. Neurosurgery. 1993; 33: 610–17.
- 25 Tatagiba M, Samii, M, Dankoweit-Timpe E, Aguiar PH, Osterwald L, Babu R, et al.. Esthesioneuroblastomas with intracranial extension. Proliferative potential and management. Arq Neuropsiquiatr. 1995; 53: 577–86.
- 26 Prevedello DM, Jagannathan J, Jane JA, Jr, Lopes MB, Laws ER, Jr. Relevance of high Ki-67 in pituitary adenomas. Case report and review of the literature. Neurosurg Focus. 2005; 19: E11.
- 27 Landolt AM, Shibata T, Kleihues P. Growth rate of human pituitary adenomas. J Neurosurg. 1987; 67: 803–06.
- 28 Morimura T, Kitz K, Stein H, Budka H. Determination of proliferative activities in human brain tumor specimens: a comparison of three methods. J Neurooncol. 1991; 10: 1–11.
- 29 Hsu DW, Hakim F, Biller BM, de la Monte SM, Zervas NT, Klibanski AH, et al. Significance of proliferating cell nuclear antigen index in predicting pituitary adenoma recurrence. J Neurosurg. 1993; 78: 753–61.
- 30 Thapar K, Kovacs K, Scheithauer BW, Stefaneanu L, Horvath E, Pernicone PJ, et al. Proliferative activity and invasiveness among pituitary adenomas and carcinomas: an analysis using the MIB-1 antibody. Neurosurgery. 1996; 38: 99–106.
- 31 Scheithauer BW, Gaffey TA, Lloyd RV, Sebo TJ, Kovacs KT, Horvath E, *et al.* Pathobiology of pituitary adenomas and carcinomas. Neurosurgery. 2006; 59: 341–53.
- 32 Preusser M, Heinzl H, Gelpi E, Schoenegger K, Harberler C, Birner P, et al. Histopathologic assessment of hot-spot microvessel density and vascular patterns in glioblastoma: poor observer agreement limits clinical utility as prognostic factors: a translational research project to the European Organization for Research and Treatment of Cancer Brain Tumor Group. Cancer. 2006; 107: 162–70.
- 33 de Lellis RA, Loyd RV, Heitz PU. Pathology and genetics: tumors of the endocrin organs (World Health Organization Classification of tumors). Lyon: International Agency for Research and Cancer; 2004.
- 34 Honegger J, Prettin C, Feuerhake F, Petrick M, Schulte-Monting J, Reincke M. Expression of Ki-67 antigen in nonfunctioning pituitary adenomas: correlation with growth velocity and invasiveness. J Neurosurg. 2003; 99: 674–79.
- 35 Mastronardi L, Guiducci A, Puzzilli F. Lack of correlation between Ki-67 labeling index and tumor size of anterior pituitary adenomas. BMC Cancer. 2001; 1: 12.
- 36 Tanaka Y, Hongo K, Tada T, Sakai K, Kakizawa Y, Kobayashi S. Growth pattern and rate in residual nonfunctioning pituitary adenomas and carcinomas: correlations

- among tumor volume doubling time, patient age, and MIB-1 index. J Neurosurg. 2003; 98: 359–65.
- 37 Jaffrain-Rea ML, di Stefano D, Minniti G, Esposito V, Bultrini A, Ferreti E, *et al.* A critical reapraisal of MIB-1 labelling index significance in a large series of pituitary tumors: secreting versus non secreting adenomas. Endocr Relat Cancer. 2002; 9: 103–13.
- 38 Yonezawa K, Tamaki N, Kokunai T. Clinical features and growth fractions of pituitary adenomas. Surg Neurol. 1997; 48: 494–500.
- 39 Paek KI, Kim SH, Song SH, Choi SW, Koh HS, Youm JY, et al. Clinical significance of Ki-67 labeling index in pituitary macroadenoma. J Korean Med Sci. 2005; 20: 489–94.
- 40 Abe T, Sanno N, Osamura YR, Matsumoto K. Proliferative potential in pituitary adenomas: measurement by monoclonal antibody MIB-1. Acta Neurochirur (Wien). 1997; 139: 613–18.
- 41 Mastronardi L, Guiducci A, Puzzilli F. Lack of correlation between Ki-67 labeling index and invasiveness among anterior pituitary adenomas: analysis of 103 cases using the MIB-1 monoclonal antibody. J Clin Pathol. 1999; 52: 107–11.
- 42 Suzuki M, Minematsu T, Oyama K, Tahara S, Miyai S, Sanno N, et al. Expression of proliferation markers in human pituitary incidentalomas. Endocr Pathol. 2006; 17: 263–75.
- 43 Delgrande E, Trouillas J, Maiter D, Donckier J, Tourniaire J. Sex-related difference in the growth of prolactinomas: a clinical and proliferation marker study. J Clin Endocrinol Metab. 1997; 82: 2102–07.
  44 Qian ZR, Li CC, Yamasaki H, Mizusawa N, Yoshimoto K,
- 44 Qian ZR, Li CC, Yamasaki H, Mizusawa N, Yoshimoto K, Yamada S, et al. Role of E-cadherin, alpha-, beta- and gamma-catenins, and p120 (cell adhesion molecules) in prolactinoma behavior. Mod Pathol. 2002; 15: 1357–65.
- 45 Wolfsberger S, Wunderer J, Zachenhofer I, Czech T, Boecher-Scwarz HG, Hainfellner J, et al. Expression of cell proliferation markers in pituitary adenomas correlation and clinical relevance of MIB-1 and anti-topoisomerase-II alpha. Acta Neurochir (Wien). 2004; 146: 831–39.
- 46 Mahta A, Haghpanah V, Lashkari A, Heshmat R, Larijani B, Tavangar SM. Non-functioning pituitary adenoma: immunohistochemical analysis of 85 cases. Folia Neuropathol. 2007; 45: 72–77.
- 47 Schaller B. Gender-related differences in growth hormonereleasing pituitary adenomas. A clinicopathological study. Pituitary. 2004; 5: 247–53.
- 48 Scheithauer BW, Kovacs KT, Laws ER, Randall RV. Pathology of invasive pituitary tumors with special reference to functional classification. J Neurosurg. 1986; 65: 733–44.
- 49 Shibuya M, Saito F, Miwa T, Davis RL, Wilson CB, Hoshino T. Histochemical study of pituitary adenomas with Ki-67 and anti DNA polymerase alpha monoclonal antibodies, bromodeoxyuridine labeling, and nucleolar organizer region counts. Acta Neuropathol (Berl). 1986; 84: 178–83.
- 50 Losa M, Franzin A, Mangili F, Terreni MR, Barzaghi R, Veglia F, et al. Proliferation index of nonfunctioning pituitary adenomas: correlation with clinical characteristics and longterm follow up results. Neurosurgery. 2000; 47: 1313–18.
- 51 Plowman PN. Pituitary adenoma radiotherapy. When, who, and how? Review. Clin Endocrinol. 1999; 51: 265–71.
- 52 Sheline GE. Pituitary tumors: radiation therapy. In: Beardwell C, Robertson GL (eds.). Clinical endocrinology. The Pituitary. London: Butterworths; 1981. pp. 106–39.
- 53 Nishioka H, Hirano A, Haraoka J, Nakajima N. Histological changes in the pituitary glands and adenomas following radiotherapy. Neuropathology. 2002; 22: 19–25.
- 54 Turner HE, Nagy Z, Gatter KC, Esiri MM, Wass JA, Harris AL. Proliferation, bcl-2, expression and angiogenesis in pituitary adenomas: relationship to tumor behavior. Br J Cancer. 2000; 14: 887–900.
- 55 Henderson WR. The pituitary adenomata. A follow-up study of the surgical results in 338 cases (Dr Harvey Cushing's series). Br J Surg. 1939; 26: 811–921.
- 56 Jefferson G. The invasive adenomas of the anterior pituitary. 2nd ed. Springfield, IL: Charles C Thomas; 1972. pp. 56–60.
- 57 Trumble HC. Pituitary tumors. Observations on large tumors which have spread widely beyond the confines of the sella turcica. Br J Surg. 1951; 39: 7–24.
- 58 Mortini P, Losa M, Barzaghi R, Boari N, Giovanelli M. Results of transphenoidal surgery in a large series of patients with pituitary adenoma. Neurosurgery. 2005; 56: 1222–33.
- 59 Jefferson G. Extrasellar extension of pituitary adenomas. Proc R Soc Med. 1940; 33: 433–58.
- 60 Trattnig S, Ba-Ssalamah A, Pinker K, Huhmann I, Wolfsberger S, Knosp E. Imaging of tumors of the pituitary gland. Wien Klin Wochenschr. 2003; 115: 23–27.

- 61 Losa M, Bazaghi RL, Mortini P, Franzin A, Mangli F, Terreni MR, et al. Determination of the proliferation and apoptotic index in adrenocorticotropin secreting pituitary tumors: comparison between micro- and macroadenomas. Am J Pathol. 2000; 156: 245–51.
- 62 Iuchi S, Saeki N, Uchino Y, Higuchi Y, Tatsuno I, Nakamura S, *et al.* Cavernous sinus invasion and tumor proliferative potential of growth hormone producing pituitary tumors. Endocr J. 2000; 47 (Suppl.): S77–79.
- 63 Iuchi T, Saeki N, Uchino Y, Osato K, Yamaura A. Proliferation, vascular endothelial growth factor expression and cavernous sinus invasion in growth homone secreting pituitary adenomas. Acta Neurochir (Wien). 2000; 142: 1345– 51.
- 64 Saeger W, Lüdecke B, Lüdecke DK. Clinical tumor growth and comparison with proliferation markers in non-functioning (inactive) pituitary adenomas. Exp Clin Endocrinol Diabetes. 2008: 116: 80–85.
- 65 Pan LX, Chen ZP, Liu YS, Zhao JH. Magnetic resonance imaging and biological markers in pituitary adenomas with invasion of the cavernous sinus space. J Neurooncol. 2005; 74: 71–76.
- 66 Pizarro CB, Oliveira MC, Coutinho LB, Ferreira NP. Measurement of Ki-67 antigen in 159 pituitary adenomas using MIB-1 monoclonal antibody. Braz J Med Biol Res. 2004; 37: 235–43.
- 67 Schreiber S, Saeger W, Lüdecke DK. Proliferation markers in different types of clinically non-secreting pituitary adenomas. Pituitary. 1999; 1: 213–20.
- 68 Wolfsberger S, Kitz K, Wunderer J, Czech T, Boecher-Schwarz HG, Hainfellner JA, et al. Multiregional sampling reveals a homogeneous distribution of Ki-67 proliferation rate in pituitary adenomas. Acta Neurochir (Wien). 2004; 146: 1323–28.
- 69 Zhao D, Tomono Y, Nose T. Expression of P27kip1 and Ki-67 in pituitary adenomas: an investigation of marker of adenoma invasiveness. Acta Neurochir (Wien). 1999; 141: 187–92.
- 70 Mizoue T, Kawamoto H, Arita K, Kurisu K, Tominaga A, Uozomi T. MIB-1 immunopositivity is associated with rapid regrowth of pituitary adenomas. Acta Neurochir (Wien). 1997; 139: 426–31.
- 71 Lath R, Chacko G, Chandy MJ. Determination of Ki-67 labeling index in pituitary adenomas using MIB-1 monoclonal antibody. Neurol India. 2001; 49: 144–47.
- 72 Nakabayashi H, Sunada I, Hara M. Immunohistochemical analysis of cell cycle-related proteins, apoptosis, and proliferation in pituitary adenomas. J Histochem Cytochem. 2001; 49: 1193–94.
- 73 Hentschel SJ, McCutcheon E, Moore W, Durity FA. P53 and MIB-1 immunohistochemistry as predictors of the clinical behavior of nonfunctioning pituitary adenomas. Can J Neurol Sci. 2003; 30: 215–19.
- 74 Ferrante E, Ferraroni M, Castrignano T, Menincatti L, Agnani M, Reimondo G, et al. Non-functioning pituitary adenoma database: a useful resource to improve the clinical management of pituitary tumors. Eur J Endocrinol. 2006; 155: 823–29.
- 75 Greenman Y, Ouaknine G, Veshchev I, Reider-Groswasser II, Segev Y, Stern N. Postoperative surveillance of clinically nonfunctioning pituitary macroadenomas: markers of tumor quiescence and regrowth. Clin Endocrinol (Oxf). 2003; 58: 763–69.
- 76 Greenman Y, Tordjamn K, Osher E, Veschchev I, Shenkerman G, Reider-Groswasser II, et al. Postoperative treatment of clinically nonfunctioning pituitary adenomas with dopamine agonists decreases tumor remnant growth. Clin Endocrinol (Oxf). 2005; 63: 39–44.
- 77 Soto-Ares G, Cortet-Rudelli C, Assaker R, Boulinguez A, Dubest C, Dewailly D, et al. MRI protocol technique in the optimal therapeutic strategy of non-functioning pituitary adenomas. Eur J Endocrinol. 2002; 146: 179–86.
- 78 Woolons AC, Hun MK, Rajapkase YR, Toomath R, Hamilton DA, Conaglen JV, et al. Non-functioning pituitary adenomas: indications for postoperative radiotherapy. Clin Endocrinol (Oxf). 2000; 53: 713–17.
- 79 Fillippella M, Galland F, Kujas M, Young J, Faggiano A, Lombardi G, et al. Pituitary tumor transforming gene (PTTG) expression correlates with the proliferative activity and recurrence status of pituitary adenomas: a clinical and immunohistochemical study. Clin Endocrinol (Oxf). 2006; 65: 536-43.

- 80 Botelho CH, Magalhães AV, Mello PA, Schmitt FC, Casulari LA. Expression of p53, Ki-67 and c-erb B2 in growth hormone –and/or prolactin –secreting pituitary adenomas. Arq Neuropsiquiatr. 2006; 64: 60–66.
- 81 Niveiro M, Aranda FI, Paya A, Boix E, Peiro G, Pico A. Oncocytic transformation in pituitary adenomas: Immunohistochemical analyses of 65 cases. Arch Pathol Lab Med. 2004; 128: 776–880.
- 82 Suliman M, Royds J, Cullen D, Timperly W, Powell T, Battersby R, *et al.* Mdm2 and P53 pathway in human pituitary adenomas. Clin Endocrinol (Oxf). 2001; 54: 317–25.
- 83 Wierinckx A, Auger C, Devauchelle P, Reynaud A, Chevallier P, Jan M, *et al.* A diagnostic marker set for invasion, proliferation, and aggressiveness of prolactin pituitary tumors. Endocr Relat Cancer. 2007; 14: 887–900.
- 84 Katznelson L, Bogan JS, Trob JR, Schoenfeld DA, Hedley-Whyte T, Hsu DW, *et al.* Biochemical assessment of Cushing's disease in patients with corticotroph macroadenomas. J Clin Endocrinol Metab. 1998; 83: 1619–23.
- 85 Korbonits M, Chahal HS, Kaltsas G, Jordan S, Urmanova Y, Khalimova Z, et al. Expression of phosphorylated p27 (Kip1) protein and Jun activation domain- binding protein 1 in human pituitary tumors. J Clin Endocrinol Metab. 2002; 87: 2635–43.
- 86 Thapar K, Kovacs K, Stefaneau L, Scheithauer B, Killinger DW, Lioyd RV, et al. Overexpression of the growth hormone releasing hormone gene in acromegaly associated pituitary tumors. An event associated with neoplastic progression. Am J Pathol. 1997; 151: 769–84.
- 87 Dubois S, Guyetant S, Menei P, Rodien P, Illouz F, Vielle B, et al. Relevance of KI-67 and prognostic factors for recurrence/progression of gonadotropic adenomas after first surgery. Eur J Endocrinol. 2007; 157: 141–47.
- 88 Comtois R, Beauregard H, Somma M, Serri O, Ris-Jilwan N, Hardy J. The clinical and endocrine outcome of transsphenoidal microsurgery of nonsecreting pituitary adenomas. Cancer. 1991; 68: 860–66.
- 89 Dekkers OM, Pereira AM, Roelfsema F, Voormolen JH, Neelis KJ, Schroijen MA, et al. Observation alone after transsphenoidal surgery for nonfunctioning pituitary macroadenoma. J Clin Endocrinol Metab. 2006; 91: 1796–801.
- 90 Drange MR, Fram NR, Herman-Bonert V, Melmed S. Pituitary tumor registry: a novel clinical resource. J Clin Endocrinol Metab. 2000; 85: 168–74.
- 91 Honegger J, Ernemann U, Psaras T, Will B. Objective criteria for successful transphenoidal removal of suprasellar nonfunctioning pituitary adenomas. A prospective study. Acta Neurochir (Wien). 2007; 149: 21–29.
- 92 Kurosaki M, Lüdecke DM, Flitsch J, Saeger W. Surgical treatment of clinically nonsecreting pituitary adenomas in elderly patients. Neurosurgery. 2000; 47: 843–48.
- 93 Lillehei KO, Kirschman DL, Kleinschmidt-Masters BK, Ridgway EC. Reassessment of the role of radiation therapy in the treatment of endocrine inactive pituitary macroadenomas. Neurosurgery. 1998; 43: 432–38.
- 94 Turner HE, Stratton IM, Byrne JV, Adams CB, Wass JA. Audit of selected patients with nonfunctioning pituitary adenomas treated without irradiation a follow-up study. Clin Endocrinol (Oxf). 1999; 51: 281–84.
- 95 Yamada S, Ohyama K, Taguchi M, Takeshita A, Morita K, Sano T. A study of the correlation between morphological findings and biological activities in clinically nonfunctioning pituitary adenomas. Neurosurgery. 2007; 61: 580–84.
- 96 Kim JH, Seo JS, Lee BW, Lee SY, Jeon SH, Lee KB. The characteristics of incidental pituitary microadenomas in 120 Korean forensic autopsy cases. J Korean Med Sci. 2007; 22 (Suppl.): S61–65.
- 97 Levy A. Pathogenesis of pituitary adenomas. In: Powell MP, Lightman SL, Laws ER, Jr (eds.). Management of pituitary tumors. The clinician's practical guide. 2nd ed. Chapter 1. Totowa, NJ: Human Press; 2003. pp. 1–20.
- 98 Romero F, Multon MC, Ramos-Morales, Dominguez A, Bernal JA, Pintor-Toro JA, et al. Human securin, hPTTG, is associated with Ku heterodimer, the regulatory subunit of the DNA-dependent protein kinase. Nucleic Acids Res. 2001; 29:
- 99 You R, Ren SG, Horwitz GA, Wang Z, Melmed S. Pituitary tumor transforming gene (PTTG) regulates placental JEG-3 cell division and survival: evidence from live cell imaging. Mol Endocrinol. 2000; 14: 1137–46.
- 100 Pei L, Melmed S. Isolation and characterization of a pituitary tumor transforming gene (PTTG). Mol Endocrinol. 1997; 11: 433–41.

- 101 Zhang X, Horowitz GA, Heaney AP, Nakashima M, Prezant TR, Bronstein MD, et al. Pituitary tumor transforming gene (PTTG) expression in pituitary adenomas. J Clin Endocrinol Metab. 1999; 84: 761–67.
- 102 Thapar K, Scheithauer BW, Kovacs K, Pernicone PJ, Laws ER, Jr. P53v expression in pituitary adenomas and carcinomas: correlation with invasiveness and tumor growth fractions. Neurosurgery. 1996; 38: 765–70.
- 103 Suhardja A, Kovacs K, Rutka J. Genetic basis of pituitary adenoma invasiveness: a review. J Neurooncol. 2001; 52: 195– 204
- 104 Ferreira JE, de Mello PA, de Magalhães AV, Botelho CH, Naves LA, Nosé V, et al. Non-functioning pituitary

- adenomas: clinical features and immunohistochemistry. Arq Neuropsiquiatr. 2005; 63: 1070–1078.
- 105 Liu W, Kunishio K, Matsumoto Y, Okada M, Nagao S. Matrix metalloproteinase-2 expression correlates with cavernous sinus invasion in pituitary adenomas. J Clin Neurosci. 2005; 12: 791–94.
- 106 Gong J, Zhao Y, Abdel-Fattah R, Amos S, Xiao A, Lopes MB, et al. Matrix metalloproteinase 9, a potential biological marker in invasive pituitary adenomas. Pituitary. 2008; 11: 37–48.
- in invasive pituitary adenomas. Pituitary. 2008; 11: 37–48.

  107 Widhalm G, Worfsberger S, Preusser M, Fischer I, Woehrer A, Wunderer J, et al. Residual nonfunctional pituitary adenomas: prognostic value of MIB labeling index for tumor progression. J Neurosurg. 2009; 111: 563–71.